Vol.5 (2015) No. 2 ISSN: 2088-5334

Preliminary Identification of Inhibition Activities towards *Eschericia coli* and *Salmonella spp.* by Pickle's Indigenous Halotolerant Bacteria

Gemilang Lara Utama[#], Fannisa Putri[#], Hanna Indah[#], Roostita Balia^{*}

Abstract— Research objectives was to determine inhibition activities towards $E.\ coli$ and $Salmonella\ spp.$ by pickle's indigenous halotolerant bacteria. Experimental methods used in the research and data were analyzed descriptive-exploratory. In order to determine pickle's indigenous halotolerant bacteria, three isolates cultured on Nutrient Agar (NA) with addition of 5 and 10% (w/v) NaCl then the total bacteria population counted. Escherichia coli and Salmonella spp. inhibition activities determined by diffusion well method on NA. Results showed that isolate P3 shown halotolerant characteristic with total bacteria population of 5.3 x 10^5 cfu/g on 5% NaCl then decreased into 5.2 x 10^3 cfu/g on 10% NaCl with no inhibition clear zone towards E.coli and 7.5 mm inhibition clear zone towards $Salmonella\ spp.$

Keywords—Pickle, Halotolerant Bacteria, Inhibition, E.coli, Salmonella spp.

I. INTRODUCTION

Preservation of vegetables can be done by fermentation, direct acidification, or combination of it with other process and additives [1]. Cucumber, eggplant, or even watermelon were fermented in salt solution to resulting in pickle.

Pickles could produced by two ways. First are the brine process which involving natural lactic acid bacteria fermentation, and the second are the fresh-pack processes without involving fermentation processes [2]. Commercial fermented cucumber mainly done with the addition of brine solution containing 6-8% NaCl [3].

Both methods using saline solution which can decrease the water activity where only certain bacteria with halotolerant abilities will grow [4]. Salting process in the making of pickle, can be role as inhibitor of undesired microoganisms growth [5].

However, the growth of pathogenic bacteria still found in pickling condition [4],[6],[7]. *Eschericia coli* found in apple cider and also *Salmonella spp*. in orange juice which the condition were similar to many pickle products [8]. Contamination risks of *E.coli* and *Salmonella spp*. in high salinity products like pickle still occur.

Inhibition abilities towards food borne pathogenic bacteria also found in bacteria isolated from home-made

fermented vegetables products [9]. The growth of pathogenic bacteria such as *E.coli* and *Salmonella spp.* can inhibited by the bacteria that involve in pickle fermentation processes through their metabolites. Generally, bacteria isolated from spontaneous pickle fermentation are *Lactobacillus plantarum*, *L. brevis*, *Leuconostoc mesenteroides*, *Pediococcus pentosaceus* and *Enterococcus faecalis* [10],[11],[12]. The bacterias produce metabolites including organic acids, carbondioxide, hydrogen peroxide, ethanol and others which role as inhibitor for pathogenic bacteria [13].

The research aims to determine pickle's indigenous halotolerant bacteria inhibition activities towards *E.coli* and *Salmonella spp.*. Potential isolates could be used to decrease contamination of *E.coli* and *Salmonella spp.* in pickle or salted products.

II. MATERIALS AND METHODS

Commercial pickle (Figure 1) bought from market, 1g sample diluted on 9 ml of 0.85% NaCl then cultured on Nutrient Agar (NA) at 37°C for 24h. Separate colony of three different isolates taken with osse and cultured on NA with addition of 5 and 10% (w/v) NaCl then the total bacteria population counted [14]. *Escherichia coli* and *Salmonella spp.* inhibition activities determined by diffusion

[#] Faculty of Agro-Industrial Technology, University of Padjadjaran, Jl. Raya Bandung Sumedang KM.21, Sumedang, 45363, Indonesia E-mail: lugemilang@gmail.com

^{*} Faculty of Animal Husbandry, University of Padjadjaran, Jl. Raya Bandung Sumedang KM.21, Sumedang, 45363, Indonesia

well method on NA [15]. Selected halotolerant bacteria, *E. coli* and *Salmonella spp.* each cultured into 15 mL Nutrient Broth (NB) and incubated for 24h at 37°C. Sweep aseptically the incubated NB of *E. coli* and *Salmonella spp.* into NA plates until covered using sterile swabs. Plug NA plates aseptically, use sterile forceps or needle to carefully pick up the plug then place 0.1 mL of incubated NB of selected isolates. Incubate the NA at 37°C for 24h then diameter of clear zones were measured.



Fig. 1. Commercial Pickle

III. RESULTS AND DISCUSSIONS

The results of preliminary characterization of the isolates shown in Table 1. Three different isolates characterized. Isolates P1 was reddish cream, cocci shape and gram negatives. Isolates P2 was cream, cocci shape and gram negatives. While isolates P3 was cream, rod shape and gram negatives.

Natural succession of microbial population in cucumber fermentations begin with high levels of aerobic bacteria, sometimes including pathogenic and spoilage organisms [16]. Lactobacillus and Leuconostoc were dominating the early stage fermentation, and Pediococcus dominated the late stage of pickle fermentation [1]. Bacteria such as L. plantarum and Leuconostoc mesenteroides was the main microorgaisms that responsible for the natural fermentation of vegetables, but L.paracasei, L. delbrueckii and L.brevis also reported as well [13]. Three bacillus strain grow well at pickling condition such as wide range of pH (3.5-10.0), temperature (15-50°C) and 4-7% NaCl [17].

TABLE I CHARACTERISTICS OF ISOLATES

Isolate	Morphol	Microscopic		
	Colour	Shape	Shape	Gram
P1	reddish cream	round	cocci	-
P2	cream	round	cocci	-
P3	cream	round	rod	-

Table 2 shown the results of halotolerant bacteria identification. Only isolates P1 that survived at 5-10% salinity with the average population of 5.3×10^5 cfu/g on 5% NaCl then decreased into 5.2×10^3 cfu/g on 10% NaCl. Other isolates did not show their ability in tolerate salinity

concentration which shown by less than 30 colonies on every plates so that can not be counted.

Halotolerant bacteria categorized based on their abilities to tolerate different salinity concentrations [14]. Bacteria with halotolerant characteristics survived in 1-5% salinity, moderately halotolerant bacteria survived in 5-20% salinity [7]. Based on the results can conclude that isolates P1 was moderately halotolerant bacteria.

TABLE II
IDENTIFICATION OF ISOLATES HALOTOLERANT CHARACTERISTICS

Isolate	5% NaCl (x10 ⁴ cfu/g)			10% NaCl (x10 ² cfu/g)		
	R1	R2	R3	R1	R2	R3
P1	53	55	52	51	50	55
P2	<30	<30	<30	<30	<30	<30
P3	<30	<30	<30	<30	<30	<30

The results of inhibition abilities towards *E.coli* and *Salmonella spp*. shown on Table 3. Table 3 showed that isolates P1 did not have inhibition abilities towards *E.coli*, but shown 7.5 mm average clear zone that indicating inhibition abilities towards *Salmonella spp*. (Figure 2). Two strain of *L. plantarum* and *P. Pentosaceus* isolated from pickle shown their abilities in inhibiting *Salmonella* invasion in animal metabolism [18].

TABLE III
INHIBITION ABILITIES TOWARDS PATHOGENS

Isolate	E. coli			Salmonella spp.		
	(mm)			(mm)		
	R1	R2	R3	R1	R2	R3
P1	-	-	-	5	10	7.5

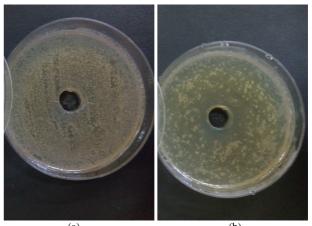


Fig. 2. Inhibition abilities (a) Isolates P1-*E.coli*, (b) Isolates P1 - *Salmonella spp*.

Pathogenic gram negatives bacteria such as *E.coli* and *Salmonella spp*. could inhibited by the production of organic acid and hydrogen proxide [19]. Lactic acid bacteria as main microflora that involves in pickle fermentation, has abilities to produce antimicrobial compound such as formic acid, benzoic acid, hydrogen peroxide, diacetyl, acetoin and bacteriocin such as nicin [20]. However, *E.coli* were more

resistant to the effects of acid than *Salmonella spp*. under typical pickling condition [21].

Bacterias isolated from pickles such as *L.plantarum*, *B. acidicola*, *B. amyloliquifaciens* and *B. mycoides*, could inhibited various food borne pathogens through production of organic acids and bacteriocins, also survived in 4-11% salinity [17]. Bacteria with halotolerant characteristics and pathogen inhibition has wide opportunities in food production. Application in salted or fermented products could decrease the contamination risk of pathogenic bacteria especially *E.coli* and *Salmonella spp.*.

IV. CONCLUSIONS

Isolate P3 shown halotolerant characteristic with total bacteria population of 5.3×10^5 cfu/g on 5% NaCl and 5.2×10^3 cfu/g on 10% NaCl with no inhibition clear zone towards *E.coli* and 7.5 mm inhibition clear zone towards *Salmonella spp*.

ACKNOWLEDGMENT

We would like to thank Laboratory of Food Microbiology, Department of Food Technology, Faculty of Agro-Industrial Technology Universitas Padjadjaran for wide range of support on this research.

REFERENCES

- Tamang, J. P. and Kailasapathy, K. Fermented foods and beverages of the world. CRC Press. 2010.
- [2] Dalmasso, J.P., "Desirable microbial growth in foods: pickles fermentation. Institut of Food Technology, Ohio State University. 2011
- [3] Doan, T., Babu, D. and Buescher, R. "Inhibition yeast in commercial pickle brines". *Journal of Food Research*, vol. 1 no. 3, 2012.
- [4] Pundir, R.K. and P. Jain, "Change in microflora of sauerkraut during fermentation and storage". World J. Dairy & Food Science, vol. p.5221-225, 2010.
- [5] Gao, S., Sun, Z., Du, X., Mao, C. And He, G. "Effect of inoculating lactic acid bacteria starter in los-walt pickle process of Zhacai". *Advace Journal of Food Science and Technology*, vol. 4, no. 6, p. 442-444, 2012.
- [6] Doyle, M.P., Beuchat, L.R. and T.J. Montville. Microbiology: fundamentals and frontiers, 2nd Ed. ASM Press, Washington, D.C. 2001
- [7] Lee, S.Y. "Microbial safety of pickled fruits and vegetables and hurdle technology". *Journal of Food Safety*, vol. 4, p. 21-32. 2004.

- [8] Breidt, F. JR., Hayes, J.S. and McFeeters, R.F. "Independent effects of acetic acid and pH on survival of *Eschericia coli* in simulated acidified pickle products". *Journal of Food Protection*, vol. 67, no.1, p.12-18, 2004.
- [9] Kazemipoor, M., Radzi, C., Begum, K., Yaze, I."Screening of antibacterial activity of lactic acid bacteria isolated from fermented vegetables against food borne pathogens". Archives des Sciences, vol. 65, no. 6. 2012.
- [10] Jagannath, A., Raju, P.S., Bawa, A.S. "A two-step controlled lactic fermentation of cabbage for improved chemical and microbiological qualities". *J. Food Qual.* vol 35 no. 1 p.13-20, 2012.
- [11] Kabak, B. and Dobson, A.D.W. "An introduction to the traditional fermented foods and beverages of Turkey". Critical Reviews in *Food Science and Nutrition*, vol. 51 p248-260. 2011.
- [12] Gardner, N. J., Savard, T., Obermeier, P., Caldwell, G., Chamagne, C.P. "Selection and characterization of mixed starter cultures for lactic acid fermentation of carrot, cabbage, beet and onion vegetable mixtures". *Int. J. Food Microbiol.*, vol. 64 no.3 p.261-75, 2001.
- [13] Irkin, R. and Songun, G.E. "Application of probiotic bacteria to the vegetable pickle products". Sci. Revs. Chem. Commun., vol. 2 no. 4, 2012.
- [14] Roohi, A., Ahmed, I., Iqbal, M., Jamil, M. "Preliminary isolation and characterization of halotolerant and halophilic bacteria from salt mines of Karak, Pakistan". Pak. J. Bot., vol. 44, p. 365-370. 2012.
- [15] Roostita, L.B., Fleet, G.H., Wendry, S.P., Apon, Z.M., and Gemilang, L.U."Determination of yeasts antimicrobial activity in milk and meat products". Advance Journal of Food Science and Technology, vol. 3 no. 6, p. 442-445, 2011.
- [16] Breidt, F. "Safety of minimally processed, acidified and fermented vegetable products". In *Microbiology of Fruits and Vegetables*, Eds. Sapers, G. M., Gorny, J. R. and Yousef, A.E., p.313-335. Boca Raton, FL:CRC Press, Inc. 2006.
- [17] Enan, G., Mahmoud, E. F., Abdel-Halie, and Tartour, E. "Evaluation of the antimicrobial activity, starter capability and technological properties of some probiotic bacteria isolated from egyptian pickles", *Life Science Journal*, vol. 11 no. 11, 2014.
- [18] Chiu, H. H., Tsai, C.C., Hsih, H.Y., Tsen, H.Y. "Screening from pickled vegetables the potential probiotic strains of lactic acid bacteria able to inhibit the *Salmonella* invasion in mice". *Journal of Applied Microbiology*, vol. 104 no. 2, p.605-612, 2008.
- [19] Ito, A., Sato, Y., Kudo, S., Sato, S., Nakajima, H. Toba, T. "The screening of hydrogen peroxide-producing lactic acid bacteria and their application to inactivating psychrotropic food borne pathogens". *Current Microbiology*, vol 47, p. 231-236, 2003.
- [20] Ozer, B., Kirmaci, H. A., Senel, E., Atamer, M., and Hayaloglu, A. "Improving the viability of *Bifidobacterium bifidum* BB-12 and *Lactobacillus acidophilus* LA-5 in white-brined cheese by microencapsulation". *International Dairy Journal*, vol 19, p. 22-29. 2009.
- [21] Lu, H. J., Breidt, JR., F., Perez-Diaz, I.M., Osborne, J. A. "Antimicrobial effects of weak acids on the survival of *Eschericia coli* O157:H7 under anaerobic conditions". *Journal of Food Protection*, vol. 74 no. 6, p.893-898, 2011.