# The Reproduction of Endemic Fish Opudi (Telmatherina prognatha Kottelat, 1991) in Lake Matano, South Sulawesi, Indonesia 

Andi Chadijah ${ }^{\text {a, }}$, , Sulistiono ${ }^{\text {b }}$, Ridwan Affandi ${ }^{\text {b }}$, Gadis Sri Haryani ${ }^{\text {c }}$, M. Ali Mashar ${ }^{\text {b }}$<br>${ }^{a}$ Program Study Aquaculture, Agricultural Faculty. Muhammadiyah Makassar University, South Sulawesi, Indonesia<br>${ }^{b}$ Department of Aquatic Resources Management, Faculty of Fisheries and Marine Sciences, Bogor Agricultural University, Bogor, Indonesia<br>${ }^{c}$ Research Centre for Limnology - the Indonesian Institute of Science, Cibinong, Jakarta, 16911, Indonesia<br>Corresponding author: *andi.chadijah@unismuh.ac.id


#### Abstract

Telmatherina prognatha is an endemic fish of the Matano Lake, in Indonesia, which demands a very crucial reproductive study, especially based on habitat conservation. This research, therefore, aims at analyzing its reproduction function as basic information for resource management in Matano Lake. Furthermore, the investigation was conducted from March 2018 to February 2019 in six sampling spots, where the habitat conditions, including the physicochemical characteristics of the water, were observed monthly. Also, the fish were sampled using a multifilament net that was 0.5 inches in mesh size and subsequently measured for total length, body weight, and gonad maturity. Moreover, the total number of specimens were approximately 1809 individuals ( n ), encompassing both sexes of male (1059) and female (750), which possessed a total length in the ranges of $\mathbf{3 6 . 4 6 - 7 2 . 0 0} \mathbf{~ m m}$ and 32.93 66.20 mm , respectively. However, the sex ratio obtained was 1.4: 1, and those with matured and maturing gonads were found in each sampling's month, and most of all, including those with spent gonads, are commonly discovered in February, March, and April. Also, the $50 \%$ probability of maturity was estimated as 57.31 mm for males and $\mathbf{4 5 . 1 6 \mathrm { mm }}$ for females, while the mean gonadal somatic indices (GSI) were $1.72 \pm 1.39$ and $4.12 \pm 1.74$, respectively. Therefore, the fecundity of this species was estimated to be in a range of $77-$ 299 eggs, and the oocyte diameter was between 0.11 mm and 1.37 mm .


Keywords-Endemic fish; reproductive; Telmatehrina prognatha; Lake Matano.
Manuscript received 25 Nov. 2019; revised 15 Dec. 2020; accepted 14 Feb. 2021. Date of publication 30 Apr. 2022. IJASEIT is licensed under a Creative Commons Attribution-Share Alike 4.0 International License

## I. Introduction

There is a high variation of endemic fish species in the ancient Matano Lake, situated around the complex of Malili Lake, and one of which is the Telmatherinidae family [1]. These water bodies possess a different height, which serves as a barrier for migrating organisms, especially from downstream to upstream. This brings about a unique organism distributing pattern with personalized endemic fish species, although the locations are close to one another [2].

The Telmatherina prognatha fish is classified in the family of Telmatherimidae, an ornamental species possessing beautiful colors. Also, it has secondary sexual characters, including different colors, shapes, or sizes, indicating dimorphism and dichromatism. This family, in general, shows mating behaviors, including (1) non-parental care, (2) spawning on substrates, (3) not exhibiting territoriality, and (4) often changing reproduction partners [3], and each species acquires different behaviors during spawning sessions.

Furthermore, some behaviors in pairs are always shown by males and females include swimming near the bottom of the water column. However, some were occasionally seen in the middle of the water column. This is a tactic of avoiding interference from other males and practiced in seeking a proper spawning ground [3]. A research conducted by Nilawati [4] reported that the specie of Telmatherina sarasinorum prefers places shaded by trees within the surrounding of the lake. Moreover, they also spread across the littoral areas, either rocky or dense riparian habitat at the edge of the water body [5].
T. prognatha is included as an extinct fish according to IUCN [6]. Therefore, investigation about its reproduction is essential, enabling its use as baseline information, concerning habitat conservation. The related studies are highly limited to some species, including T. celebensis [7]-[9], T. antoniae [10] and T. sarasinorum [4], [11]. This research aims to analyze the reproduction of $T$. prognatha, later to serve as a
fundamental source of information on fish resource management in the Matano Lake.

## II. Materials and Method

## A. Material

This study was conducted in the Matano Lake, Southeast Sulawesi, for 12 months, i.e., from March 2018 to February 2019, and the sampling locations composed of six stations, which include Lawa River, Wotu Pali, Salonsa Beach, Utuno, Petea River, and Tanah Merah (Figure 1).


Fig. 1 Research location in the Matano Lake

## B. Methods

Sampling fish was carried out once every month directly using a multifilament net in size of 30 m length, 2 m width, and 0.5 -inch mesh size. The net was stretched on the upper column near-surface water shaping an angle range of $45^{\circ}-90^{\circ}$ perpendicularly on the coastline in each station. The sampled fish were measured and weighted using calipers and an analytical scale with an accuracy of 0.01 mm and 0.001 g , respectively. These fish were then dissected, and their sexes were defined. Their gonads maturity development was divided into some levels [4], [12].


Fig. 2 Sampled fish of Telmatherina prognatha male (a) and female (b) Kottelat 1991

The determination of gonad maturity was assessed by morphologically observing shape, size, color, and gonads’ content development. Hence they were subsequently classified into some stages of maturation [4], [12].

## C. Data Analysis

1) Sex Ratio: This sex ratio analysis used the following formula.

$$
\begin{equation*}
X=\frac{\mathrm{M}}{\mathrm{~F}} \tag{1}
\end{equation*}
$$

Where:
X = Sex ratio
$\mathrm{M}=$ Number of male fish (individuals)
$\mathrm{F}=$ number of female fish (individuals)
Furthermore, a chi-square test was taken, using the formula [13] as follows.

$$
\begin{equation*}
X^{2}=\frac{(O i j-E i j)^{2}}{E i j} \tag{2}
\end{equation*}
$$

Where:
$\mathrm{X}^{2}=$ value of chi-square
$\mathrm{Oij}=$ frequency number of catches
$\mathrm{Eij}=$ expected frequency of occurrence
2) Fish Sizes of First Gonad Maturity: Supposing the first gonad maturity (Lm50) fish sizes was based on the total length of sampled fish applying the following formula [14].

$$
\begin{equation*}
P=\frac{1}{\left[1+e^{-r(L-L m)}\right]} \tag{3}
\end{equation*}
$$

Where:
$P=$ probability of fish with gonad maturity (\%)
$\mathrm{e}=$ exponential of natural number
$\mathrm{L}=$ average length (mm)
$\mathrm{Lm}=$ size of first gonad maturity $(\mathrm{mm})$
3) Gonadal Somatic Index: The Gonadosomatic Index (GSI) refers to the following formula [15].

$$
\begin{equation*}
\mathrm{GSI}=\frac{\mathrm{GW}}{\mathrm{BW}} \times 100 \tag{4}
\end{equation*}
$$

Where:
GMI= gonado somantic index.
$\mathrm{GW}=$ gonad weight $(\mathrm{g})$
$\mathrm{BW}=$ body weight (g)
4) Fecundity: Fecundity was calculated by applying a direct count method on fish eggs, one after the other. Therefore, its relationship with fish body length is involved using the formula [16] as follows.

$$
\begin{equation*}
\mathrm{F}=\mathrm{aLb} \tag{5}
\end{equation*}
$$

Where:
$\mathrm{F}=$ number of eggs (item)
$\mathrm{L}=$ Total length (mm)
a and $\mathrm{b}=$ constants

## III. Results and Discussion

## A. Sex Ratio

During a sampling period, the total number of captured $T$. Prognatha, starting from March 2018 to February 2019, was about 1,809 individuals, consisting of $58.54 \%$ male and $41.46 \%$ female. Furthermore, the sex ratio utilizing the Chi-Square test with a significant level of 0.05 indicates an imbalance between both, at 1.41: 1 , in which the $\mathrm{X}^{2}$ table $>\mathrm{X}^{2}$ count.

TABLE I
The sex ratio of fish T. prognatha spatially

| Station | Male |  | Female |  | Sex | $\mathbf{X}^{\mathbf{2}}$ |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Individual | $\mathbf{( \% )}$ | Individual | (\%) | Ratio | count |
| S. Lawa | 175 | 16.53 | 102 | 13.60 | $1.72: 1$ | $19.24^{*}$ |
| Wotu | 100 | 9.44 | 63 | 8.40 | $1.59: 1$ | $8.40^{*}$ |


| P. | 241 | 22.76 | 157 | 20.93 | $1.54: 1$ | $17.73^{*}$ |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| Salonsa | 214 | 20.21 | 180 | 24.00 | $1.19: 1$ | 2.93 |
| Utuno | 109 | 10.29 | 108 | 14.40 | $1.01: 1$ | 0.005 |
| S. Petea | 220 | 20.77 | 140 | 18.67 | $1.57: 1$ | $17.78^{*}$ |
| Tanah <br> Merah | Where: $\chi^{2}$ table $(0.05,1)=3.84 ; \chi^{2}$ table $\geq \chi^{2}$ count. A comparison of male and <br> female $=1: 1 ;(*)$ the comparison of male and female $\neq 1: 1$ |  |  |  |  |  |

Spatially, the ratio in each location possessed different values, and the females were always lesser, comparatively. However, the sampling point with the highest male was Lawa River (1.72:1), and two others recorded 1:1, including Utuno and Petea River (Table 1).

TABLE II
The sex ratio of fish $T$. prognatha temporally

| Month | Male |  | Female |  | $\begin{gathered} \text { Sex } \\ \text { Ratio } \end{gathered}$ | $\begin{gathered} \mathrm{X}^{2} \\ \text { count } \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Individual | (\%) | Individual | (\%) |  |  |
| March'18 | 88 | 8.31 | 75 | 10.00 | 1.17:1 | 1.04 |
| April' 18 | 95 | 8.97 | 63 | 8.40 | 1.15:1 | $6.48{ }^{*}$ |
| May' 18 | 58 | 5.48 | 51 | 6.80 | 1.14:1 | 0.45 |
| Jun'18 | 59 | 5.57 | 41 | 5.47 | 1.44:1 | 3.24 |
| Jul'18 | 57 | 5.38 | 43 | 5.73 | 1.33:1 | 1.96 |
| Aug'18 | 126 | 11.90 | 97 | 12.93 | 1.30:1 | 3.77 |
| Sept' 18 | 195 | 18.41 | 94 | 12.53 | 2.07:1 | $35.30{ }^{*}$ |
| Oct'18 | 89 | 8.40 | 88 | 11.73 | 1.01:1 | 0.01 |
| Nov'18 | 151 | 14.26 | 89 | 11.87 | 1.70:1 | 16.02* |
| Dec'18 | 52 | 4.91 | 36 | 4.80 | 1.44:1 | 2.91 |
| Jan'19 | 50 | 4.72 | 43 | 5.73 | 1.16:1 | 0.53 |
| Feb'19 | 39 | 3.68 | 30 | 4.00 | 1.30:1 | 1.17 |

Where: $\chi^{2}$ table $(0.05,1)=3.84 ; \chi^{2}$ table $\geq \chi^{2}$ count. A camparison of male and female $=1: 1 ;\left(^{*}\right)$ the comparison of male and female $\neq 1: 1$

Temporally, the ratio during sampling periods was balanced, and the conditions where the ratio was $\neq 1: 1$ were only detected in three months, encompassing April, September, and November. However, the highest and the lowest occurred in September (2.07:1) and October (1.01:1), respectively. Even though the sampling period, the male was dominantly caught than the females, although the Chi-Square test towards the species monthly resulted in a balanced proportion (Table 2).

Commonly, the sex ratio of $T$. prognatha was $1: 1$, except those found in the Lawa River station, which was $\neq 1: 1$. This ratio was probably affected by several natural and environmental factors [17]. Also, some rainbow fish that also possessed this $1: 1$ ratio to include $T$. celebensis from Towuti Lake [7], and T. antoniae [18], while T. sarasiunorum [4] varied with sampling location and period, although the highest was recorded as $3.05: 1$. This imbalanced proportion was confirmed from observation during the spawning behavior as males fought over some females [4]. According to research conducted by Jayadi et al. [20], the small cyprinids fish (Mystacoleucus padangensis) occurs at a sex ratio of 1:1.99. The opposite occurs in Celebes rainbowfish (Marostherina ledigesi), where the male is smaller than the female [20]. This finding was also reported for the marmorated ricefish (Oryzias marmoratus) [21]. Also, preserving the fish population requires that the comparison between both sexes is expected to be balanced (1:1), or having more females [22]. Sulistiono [23] also explained that some differences between both genders were due to their activity in
waters, adaptable ability, and genetic factors. Also, the movement can be influenced by the number of catch that affects the ratio [24].

## B. Gonad Maturity Stage (GMS)

According to sampling locations and time, the percentage of gonadal maturity stage of $T$. prognatha fluctuated between male and female. Spatially, the highest and the lowest for the males with the GMS I were abundantly found in Lawa and Patea River, respectively, and those with the GMS I, II, III, and IV were commonly identified in Salonsa Island and Wotu Pali, respectively. Furthermore, those with GMS V were discovered in Utuno and Petea River, respectively (Figure 3).


Fig. 3 The GMS percentage of male (left) and female (right) fish Telmatherina prognatha spatially

Conversely, the highest percentage of female fish were found in four locations, which include Wotu Pali (GMS 1), Utuno (GMS II and III), and Salonsa Island (GMS IV). Meanwhile, the lowest were discovered in two locations, encompassing Utuno (GMS II and III) and Wotu Pali (GMS IV). Also, the lowest with GMS V was only detected in three locations: Wotu Pali, Salonsa Island, and Utuno. Furthermore, a lower proportion of fish with matured gonads in Wotu Pali was presumed to be because the habitat is a nursery ground, indicating that it consists of rocky and rooty structures in the narrowly litoral area.

GMS I, III, and IV tend to dominate within each sampling period (Figure 4), with the highest male of GMS III and IV found in September and the lowest occurring in January and February. Meanwhile, the females were mostly noticed in August for GMS III, September, and December for GMS IV. However, those with spent gonad conditions (also with GMS III and IV), mainly females, were abundant in three consecutive months, including February, March, and April, implying that these were the peak season for spawning. Nevertheless, August to December also showed a high percentage compared to the remaining months of the year.


Fig. 4 The GMS percentage of male (left) and female (right) fish Telmatherina prognatha temporally

The sampling location and periods were determinants of the different percentages of fish and its GMS, where the value was highest from February to April and from August to December. Also, a similar pattern was also demonstrated by the GMI evaluations that were frequently higher in those months. Furthermore, those with gradual spawning on gonad had acquired gonadal maturity, comprising various stages with different percentages [25]. According to Nilawati [4], the amount of $T$. sarasinorum (GMS IV) in the late dry and the early wet seasons were presumed since those periods were the peak season of spawning, followed by the rise in water surface of the lake. The report of Mamangkey and Nasution [26] express that the GMS numbers of a goby fish, endemic to Sulawesi (Glossogobius matanensis), with different gonad maturity, occurred abundantly in two different depths ( 75 m and 100 m ), with a peak of spawning that took place from March to April and October to December [24].

## C. Sizes of First Maturity Gonad

The size of T. prognatha on first gonad maturity (reaching GMS IV) was 43.43 mm long and 1.4 g in weight for male, while the female spanned 39.00 mm in length and 0.82 g in weight. Therefore, the total lengths (TL) for both, with $50 \%$ probability maturing gonads, were 57.31 mm and 45.16 mm , respectively (Figure 5). Hence, revealing that at the same size, the females were comparably quicker to obtain maturity. This size is further related to their growth, environmental effects, and reproductive strategy.


Fig. 5 The sizes of male and female fish Telmatherina prognatha with probability $50 \%$ maturing gonad

The first gonadal maturity (GMS IV) size for both male and female $T$. prognatha was 47.43 mm and 39.00 mm , respectively, and the highest number was obtained in

September. Furthermore, an analysis of the results calculating $50 \%$ probability of gonadal maturity for both sexes recorded total lengths of 54.3 mm and 53.4 mm . Meanwhile, $T$. celebensis indicated a first gonadal development in the 37.3 mm long male and 36.4 mm for females [7], while the outcome of the current research demonstrates that more than $50 \%$ of the studied species were captured in the GMS III and IV. Also, a comparable result was obtained in the ricefish ( $O$. marmoratus), where GMS I-V were noticed in every month [21].

## D. Gonadosomatic Index (GSI)

The gonadal somatic index (GMI) was higher in line with the gonad maturity stage with a mean value of both sexes of fish in each location and period of sampling that signifies the male was smaller than the female at $1.72 \pm 1.39$ and $4.12 \pm 1.74$, respectively.


Fig. 6 The gonadosomantic index (GSI) of fish Telmatherina prognatha spatially

Spatially, the highest and the lowest mean of GSI for the male $T$. prognatha were located in Wotu Pali station ( $2.28 \pm 1.85$ ) and Lawa River ( $1.39 \pm 0.9672$ ), respectively. Furthermore, for the female were in Petea River (4.32 $\pm 1.63$ ) and Lawa River ( $3.88 \pm 1.77$ ), respectively (Figure 6).

The highest and lowest mean of GSI for the male fish $T$. prognatha temporally occurred in March ( $4.39 \pm 1.96$ ) and January ( $1.10 \pm 0.61$ ), respectively, while that for females was in March ( $5.37 \pm 2.04$ ) and February ( $3.06 \pm 1.28$ ), respectively (Figure 7). Figure 7 infers that the most significant record ensued in five months, including March, May, September, November, and December, indicating that spawning is higher in these periods.

The GSI values of female $T$. prognatha were generally higher, which is similar to the report by Jayadi et al. [9], with outcomes for the female $T$. celebensis being more (1.58$2.64 \%$ ) than the male ( $0.38-0.82 \%$ ). These results enable the indication of reproductive activity peaks, as the highest and lowest for the male T. prognatha were recorded in March and July, respectively, while the least for females occurred in April. However, on four months (March, May, September, November, and December), the values were observed to
commonly be higher than others, although the index fluctuates, which enables the indication that spawning occurs on time [22].


Fig. 7 The gonadosomantic index (GSI) of fish Telmatherina prognatha temporally

This finding was not similar with the study by [9], where the GSI peaks for T. celebensis in the Matano Lake happened in June and October while in Towuti Lake, it occurred in two months (November and February). Also, different spawning periods of Japenese whiting fish (Sillago japonica) appeared in two different locations of capture [27], where the highest value for both sexes was from April to August, and May, respectively [28]. This result indicates the variance of the period based on species and the ecological property of the habitats [29].

## E. Fecundity

The fecundity of $T$. prognatha was at an average of 150 (GMS IV), with the lowest ( 77 eggs) occurring int fish that has a total length of 57.20 mm , and the highest ( 299 eggs ) was from a 66.05 mm long type. Furthermore, the coefficient correlation of the regression equation test shows $r=0.16$ (Figure 8), demonstrating a relationship between fecundity and the total length of $T$. prognatha. Also, it can also be inferred that the number of eggs in the ovaries were incapable of being surmised through the length data obtained.


Fig. 8 The relationship between fecundity and the total length of fish Telmatherina prognatha

Fecundity is highly related to the environment, based on the existence of observed proportional changes [23], and the result obtained for T. prognatha was in a range of 77-299 eggs,
with the average of 178 eggs. This result is well known to be affected by age, size, species, food abundance, season, and environmental conditions [29]. Furthermore, the data obtained signify that total lengths of T. prognatha possessing the lowest and highest values were 50.58 mm and 66.05 mm , respectively. Meanwhile, [9] expressed the fecundity of $T$. celebensis in Matano Lake to be within the range of 297-1,265 and that in Towuti Lake was 185-1448 eggs [7]. Nilawati [4] conveyed $T$. sarasinorum has a range of 64-488 eggs, while that of Celebes rainbowfish Marosatherina ladigesi defers, depending on the food type eaten, although when fed with Daphnia sp., 65-281 eggs were recorded [20]. These differences may be due to the larger age class used in the calculations [17], while Jayadi et al. [20] indicated that low fecundity was presumed to be related with small body shape, as seen in the Celebes rainbowfish (M. ladigesi).

In terms of the correlation with total length, the numbers of T. prognatha eggs were not predictable, which is in line with Karyanti [30], stating that fecundity had a weak association with length and weight. Furthermore, a low $R^{2}$ value demonstrates the absence of a relationship [22], [23]. However, research conducted by Suryanti [31] indicated a positive correlation in small cyprinids fish (M. padangensis) located in Naborsahaan, where an upsurge followed the increase in length and weight in the rate of fecundity.

## IV. Conclusion

The sex ratio obtained was 1.4: 1. Size of maturity was estimated as 57.31 mm for male and 45.16 mm for female, the fecundity of this specie was estimated to be in a range of 77299 eggs, and partial spawner. The peak of spawning occurs in February-April with spawning areas around the area of P . Salonsa and Utuno.

## Acknowledgement

The authors are grateful to the Excellence Scholarship of Domestic Affairs (BUDI-DN/LPDP) for funding this research.

## REFERENCES

[1] S. B. Andy Omar. Fish World. Yogyakarta. Gadjah Mada University Press, Indonesia, 2012.
[2] A. Nontji. Natural Lakes of Nusantara. Ali F, Tanjung LR, Widiyanto T, Henny C, Ridwansyah I, Sulastri, Hidayat, Subehi L, Wibowo H, M.Sc, Maghfiroh M, editor. Indonesia (ID): Pusat Penelitian Limnologi, Lembaga Ilmu Pengetahuan Indonesia, Indonesian, 2017.
[3] S. M. Gray, J. S. McKinnon. A Comparative Description of Mating Behaviour in the Endemic Telmatherinid Fishes of Sulawesi"s Malili Lakes. Environment Biology Fishesries., vol. 75, pp. 471-482, 2006.
[4] J. Nilawati. Reproduction of fish Telmatherina sarasinorum (Kottelat, 1991) As A Basis of Conservation in the Matano Lake, Sout Sulawesi. [disertasi]. Bogor (ID): Institut Pertanian Bogor, 2012.
[5] A. Chadijah, Sulistiono, G. S. Haryani, R. Affandi, A. Mashar. Species Composition of Telmatherina Caught in The Vegetated and Rocky Habitats in Matano Lake, South Celebes, Indonesia. Aquaculture, Aquarium, Conservation \& Legislation International Journal of the Bioflux Society., vol. 11 Issue 3, pp. 948-955, 2018.
[6] [IUCN] International Union Conservation of Nature. 2008. Redlist of Threatened Species. [online]. Available at: www.redlist.org.
[7] S. H. Nasution. The reproductive characteristics of Selebensis rainbow endemic fish (Telmatherina celebensis Boulenger) in the Matano Lake. Jurnal Penelitian Perikanan Indonesia. vol. 11, pp. 29-37, 2005.
[8] S. H. Nasution, Sulistiono, D. S. Sjafei, G. S. Haryani GS. 2007. Spasial Distributioni spasial and temporal endemic fish rainbow selebensis (Telmatherina celebensis Boulenger) di Lake Towuti,

South Sulawesi. Journal Penelitian Perikanan Indonesia., vol 12, no. 2, 2007
[9] Jayadi, H. Rimal, Arifuddin. The reproduction of Sulawesi rainbow endemic fish Telmatherina celebensis in the Matano Lake, South Sulawesi. Torani (Jurnal Ilmu Kelautan dan Perikanan), vol. 20, no. 1 pp. $44-48,2010$.
[10] F. Y. Tantu, Sulistiono, I. Muchsin. Habitat, distribution, and size structure of fish opudi (Telmatherina antoniae) in the Matano Lake. Jurnal Agrisains., vol. 13, no. 2, pp. 140-148, 2012.
[11] J. Nilawati, Sulistiono, D. S. Sjafei, M. F. Rahardjo, I. Muchsin. Spawning Habitat of Telmatherina sarasinorum (Family: Telmatherinidae) in Lake Matano. Jurnal Iktiologi Indonesia., vol. 10, no. 2, pp. 101-110, 2010.
[12] N.D. Koç, Y. Aytekin, R. Yüce. Ovary maturatıon stages and histological investigation of ovary of the Zebrafish (Danio rerio). Journal Brazilian Archives of Biology and Technology., vol. 51, no. 3, pp. 513-522, 2008.
[13] Sudjana. Statistic Method $6^{\text {th }}$ Edition. Bandung: Tarsito, Indonesia, 1992.
[14] M. King. Fisheries biology, assessment and management. Fishing News Books. London, 1995.
[15] M. I. Effendie. Fishery Biology Method. Yayasan Dewi Sri, Bogor. Indonesia. 1979.
[16] C. P. Le Cren. Length-Weight relationship and seasonal cycle in gonad weight and condition in the Perch (Perca fluviatilis). Journal of Animal Ecology., vol. 20, no. 2, pp. 201-219, 1951.
[17] B. Kamangar, E. Ghaderi, H. Hoseinpour. Growth and reproductive biology of Capoeta damascina (Valenciennes, 1842) from a tributary of Tigris. Iranian Journal of Fisheries Sciences., vol. 14, no. 4 pp. 956969, 2015.
[18] F.Y. Tantu Reproductive ekobiology of fish Telmatherina antoniae (Kottelat, 1991) as a basis of conserving the endemic fish in the Matano Lake, South Sulawesi. [dissertation]. Bogor (ID): Institut Pertanian Bogor. Indonesian, 2012.
[19] A. Suryanti, Sulistiono, I. Muchsin I, E. S. Kartamihardja. Spawning habitat and nurshery ground of fish bilih Mystacoleucus padangensis (Bleeker, 1852) in the Naborsahan River, Toba Lake, North Sumatera. Jurnal Bawal., vol. 9 pp. 33-42, 2017.
[20] Jayadi, Hadijah, B. Tang, A. Husma. Reproductive biology of fish besang-besang (Marosatherina ledigasi Ahl, 1936) in some rivers in South Sulawesi. Jurnal Iktiologi Indonesia., vol 16, no. 2, pp. 185-198, 2016.
[21] Sulistiono. Reproduction of fish lunjar (Oryzias marmoratus) in the Towuti Lake, Southeast Sulawesi. Jurnal Agrisains., vol. 13, no. 1, pp. 55-65, 2012.
[22] Sulistiono. Reproduction of fish rejung (Sillago sihima Forsskal) in the Mayangan waters, Subang, Jawa Barat. Jurnal Iktiologi Indonesia., vol. 11, no. 1, pp. 55-65, 2011.
[23] Sulistiono. Reproduction of fish beloso (Glossogobius giuris) in the waters of Ujung Pangkah, East Java. Jurnal Akuakultur Indonesia., vol. 11, no.1, pp. 64-75, 2012.
[24] Sulistiono, A. Firmansyah, S. Sofiah, M. Brojo, R. Affandi, J. Mamangke. Biological aspects of fish butini (Glossogobius matantensis) in the Towuti Lake, South Sulawesi. Jurnal Ilmu-ilmu Perairan dan Perikanan Indonesia., vol. 14, no. 1, pp. 13-22, 2007.
[25] R. Cárdenas, M. Chávez, J. González, P. Aley, J. Espinosa, L.F. Jiménez-García. Oocyte structure and ultrastructure in the Mexican silverside fish Chirostoma humboldtianum (Atheriniformes: Atherinopsidae). Revista de biologia tropical. 56 pp. 1825-1835, 2008.
[26] J.J. Mamangkey, S. H. Nasution SH. Reproduction of endemic fish butini (Glossogobius matanensis Weber 1913) based on the depth and times in the Towuti Lake, South Sulawesi. Jurnal Biologi Indonesia., vol. 8, pp. 31-43, 2012.
[27] Sulistiono, S. Watanabe, M. Yokota. Reproduction of the Japanese Whiting, Sillago japonica, in Tateyama Bay. Suisanzoshoku., vol. 47, no. 3, pp. 209-214, 1999.
[28] Sulistiono, I. Wibisana, P. P. Sari, R. Affandi, S. Watanabe, M. Yokota. Maturity and food habits of the Japanese whiting (Sillago japonica) in Omura Bay. Nagasaki Japan. Jurnal Ilmu-ilmu Perairan dan Perikanan Indonesia., vol. 9, no. 2, pp. 121-128, 2002.
[29] F. Özdemir, F. Erk'akan F. Growth and reproductive properties of an endemic species, Gobio hettitorum Ladiges, 1960, in Y eşildere Stream, Karaman, Turkey. Hacettepe Journal of Biological Chemistry., vol. 40, no. 4, pp. 457-468, 2012.
[30] Karyanti, S.B. Andy Omar, J. Tresnati. Analysis of fecundity and eggs diameter of fish beseng-beseng (Marosatherina ladigesi Ahl, 1936) in Pattunuang Asue River and Bantimurung River, Maros Regency. Prosiding Simposium Nasional I Kelautan dan Perikanan, 3 Mei 2014; Makassar, Indonesia. pp. 1-10, 2014.
[31] A. Suryanti, Sulistiono, I. Muchsin I, E. S. Kartamihardja. Reproductive biology of female bilih fish Mystacoleucus padangensis (Bleeker, 1852) in Naborsahan River, Toba Lake, North Sumatera, Indonesia. Asian Journal of Developmental Biology., pp. 1-12, 2016.
[32] Azrita, H. Syandri. Fecundity, egg diameter and food Channa Lucius Cuvier in different waters habitats. Journal of Fisheries and Aquaculture., vol. 4, pp. 115-120, 2013.

